## PROTEIN SEQUENCE ANALYSIS



## Need good protein sequence analysis tools because:

- As number of sequences increases, so gap between seq data and experimental data increases
- But increase number of sequences increase sequence DB and therefore increased chance of finding similar sequence
- Computer analysis can narrow down number of functional experiments required



## **UNKNOWN PROTEIN SEQUENCE**

#### LOOK FOR:

- Similar sequences in databases ((PSI) BLAST)
- Distinctive patterns/domains associated with function
- Functionally important residues
- Secondary and tertiary structure
- Physical properties (hydrophobicity, IEP etc)



### BASIC INFORMATION COMES FROM SEQUENCE

- One sequence- can get some information eg amino acid properties
- More than one sequence- get more info on conserved residues, fold and function
- Multiple alignments of related sequences- can build up consensus sequences of known families, domains, motifs or sites.
- Sequence alignments can give information on loops, families and function from conserved regions



#### LEVEL OF FUNCTION INFORMATION IN PROTEIN SEQUENCES



## AMINO ACID PROPERTIES

- Small
- Small hydroxyl
- Basic
- Aromatic
- Small hydrophobic
- Medium hydrophobic
- Acidic/amide
- Small/polar

Ala, Gly Ser, Thr His, Lys, Arg Phe, Tyr, Trp Val, Leu, Ile Val, Leu, Ile, Met Asp, Glu, Asn, Gln Ala, Gly, Ser, Thr, Pro



#### **Protein functions from specific residues**

- C disulphide-rich, metallothionein, zinc fingers
- DE acidic proteins (unknown)
- G collagens
- H histidine-rich glycoprotein
- KR nuclear proteins, nuclear localisation
- P collagen, filaments
- SR RNA binding motifs
- ST mucins

- Polar (C,D,E,H,K,N,Q,R,S,T) active sites
- Aromatic (F,H,W,Y) protein ligandbinding sites
- Zn+-coord (C,D,E,H,N,Q) active site, zinc finger
- Ca2+-coord (D,E,N,Q) ligand-binding site
- Mg/Mn-coord (D,E,N,S,R,T) Mg2+ or Mn2+ catalysis, ligand binding
- Ph-bind (H,K,R,S,T) phosphate and sulphate binding



## **Protein functions from regions**

- Active sites- short, highly conserved regions
- Loops- charged residues and variable sequence
- Interior of protein- conservation of charged amino acids



#### **Additional analysis of protein sequences**

- transmembrane regions
- signal sequences
- localisation signals
- targeting sequences
- GPI anchors
- glycosylation sites

- hydrophobicity
- amino acid composition
- molecular weight
- solvent accessibility
- antigenicity



#### FINDING CONSERVED PATTERNS IN PROTEIN SEQUENCES

- Pattern short, simplest, but limited
- Motif conserved element of a sequence alignment, usually predictive of structural or functional region

To get more information across whole alignment:

- Matrix
- Profile
- HMM



## PATTERNS

- Small, highly conserved regions
- Shown as regular expressions Example:
  - $[AG]-x-V-x(2)-x-\{YW\}$
  - [] shows either amino acid
  - X is any amino acid
  - X(2) any amino acid in the next 2 positions
  - { } shows any amino acid except these

#### **BUT-limited to near exact match in small region**



## MATRIX

- 210 possible aa pairs (190 different aa, 20 identical aa)
- Start with sequence alignment and build up a table of probabilites of finding each aa in each position of the sequence
- Can be scored in several different ways



#### Matrix scores can be based on:

- **Genetic code** -base changes required to convert codons for 2 amino acids
- Chemical similarity -polarity, size, shape, charge
- **Observed substitutions** -based on analysing frequencies seen in alignments- inter-reliable
- **Dayhoff mutation data matrix** likelihood of mutation from one aa to another, but different positions are not equally mutatable, and only useful for close function because sequence alignments are very related proteins



## Matrix scoring continued

- **BLOSUM** -matrix from ungapped alignments of distantly related sequences -cluster sequences similar at a threshold value of % identity -substitution frequencies for all pairs of aa calculated -used to calculate a log odds BLOSUM (blocks substitution matrix). Can vary threshold values
- **3D structure matrix** -derived from tertiary structure alignment, good, but only used if structure is known **Best matrices are derived from observed substitution** data, it is important to use select scoring appropriate for evolutionary distance interested in.



## PROFILES

- Table or matrix containing comparison information for aligned sequences
- Used to find sequences similar to alignment rather than one sequence
- Contains same number of rows as positions in sequences
- Row contains score for alignment of position with each residue



#### **Example of a Profile**

ਸ	ĸ	т.	т.	g	н	C C	т.	т.	37		
ב ה	v	1	ы Т	~			11 14	ы Т			
F	л Л	A -	r 	ۍ ۳	Q 0	T	19	r -	Q		
<u>х</u>	Р	T	V 	G	Q	E	ц –	ц –	G		
F	Ρ	V	V	K	E	A	I	Г	K		
F	К	V	Г	A	A	V	I	A	D		
Г	Ε	F	I	S	Ε	С	I	I	Q		
F	К	L	$\mathbf{L}$	G	Ν	V	L	V	С		
A		-18	-10	-1	-8	8	-3	3	-10	-2	-8
С		-22	-33	-18	-18	-22	-26	22	-24	-19	-7
D		-35	0	-32	-33	-7	6	-17	-34	-31	0
Е		-27	15	-25	-26	-9	23	-9	-24	-23	-1
F		60	-30	12	14	-26	-29	-15	4	12	-29
G		-30	-20	-28	-32	28	-14	-23	-33	-27	-5
н		-13	-12	-25	-25	-16	14	-22	-22	-23	-10
I		3	-27	21	25	-29	-23	-8	33	19	-23
К		-26	25	-25	-27	-6	4	-15	-27	-26	0
г		14	-28	19	27	-27	-20	-9	33	26	-21
М		3	-15	10	14	-17	-10	-9	25	12	-11
N		-22	-6	-24	-27	1	8	-15	-24	-24	-4
Р		-30	24	-26	-28	-14	-10	-22	-24	-26	-18
Q		-32	5	-25	-26	-9	24	-16	-17	-23	- 7
R		-18	9	-22	-22	-10	0	-18	-23	-22	-4
s		-22	-8	-16	-21	11	2	-1	-24	-19	-4
т		-10	-10	-6	-7	-5	-8	2	-10	-7	-11
v		0	-25	22	25	-19	-26	6	19	16	-16
W		9	-25	-18	-19	-25	-27	-34	-20	-17	-28
Y		34	-18	-1	1	-23	-12	-19	0	0	-18

Match values are higher for conserved residues



## **Building a Profile**

- To get good profile need good, hand-curated alignment
- Use alignment to build up position-specific scoring matrix
- Use matrix (profile) to do PSI-BLAST with several iterations



## **SCORES**

 E-value is chance of a random sequence sequence hitting. E-value 1.0 not significant, 0.1 possibly significant,< 0.01 most likely to be significant. All depends on database size



#### HIDDEN MARKOV MODELS (HMM)

- An HMM is a large-scale profile with gaps, insertions and deletions allowed in the alignments, and built around probabilities
- Package used HMMER (http://hmmer.wusd.edu/)
- Start with one sequence or alignment -HMMbuild, then calibrate with HMMcalibrate, search database with HMM
- E-value- number of false matches expected with a certain score
- Assume extreme value distribution for noise, calibrate by searching random seq with HMM build up curve of noise (EVD)



## REPEATS

- Structural and evolutionary entities found in 2 or more copies
- Often assemble into elongated "rods", "superhelices" or "barrel" structures
- Specialised cases when building profiles



## PITFALLS OF METHODS

- **BLAST** only pick up homologues, not distant, divergent family members
- **PSI-BLAST** fine for superfamilies, not very good for small very conserved motifs
- **Patterns** small, localised and need to be highly conserved regions
- **HMMER** slow process for searching database
- **Profiles** if false positive picked up, pulls in its companions, in large families members can be missed
- Alignment methods automatic, less biological significance



# **Big problem in protein sequence analysis- multidomain proteins:**

- Most conserved domain will score highest in sequence similarity searches, may overlook lower scoring domains
- Iterative searching of multi-domain proteins could pick up unrelated proteins



#### **SUMMARY OF PATTERN METHODS**



## COMMON PROTEIN PATTERN DATABASES

- Prosite patterns
- Prosite profiles
- Pfam
- SMART

- Prints
- ProDom
- DOMO
- BLOCKS

## SOFTWARE FOR PROTEIN SEQUENCE ANALYSIS

- GCG (http://www.gcg.com/)
- EMBOSS (ftp:ftp.sanger.ac.uk/pub/EMBOSS)
- PIX- HGMP (http://www.hgmp.mrc.ac.uk)
- ExPASy Proteomics tools (http://www.expasy.org/tools)
- PredictProtein (http://www.emblheidelberg.de/predictprotein/)

