Microarray Data Analysis - II

FIOCRUZ Bioinformatics Workshop 6 June, 2001

Challenges in Microarray Data Analysis

- **Spot Identification and Quantitation.**
- Normalization of data from each experiment.
- **Identification of Differentially Expressed Genes**
- Identified of genes with correlated patterns of expression.
- Interpretation of data with respect to pathways.
- **Literature filtered analysis.**

Image Processing Issues

Spot Finding

Background Subtraction

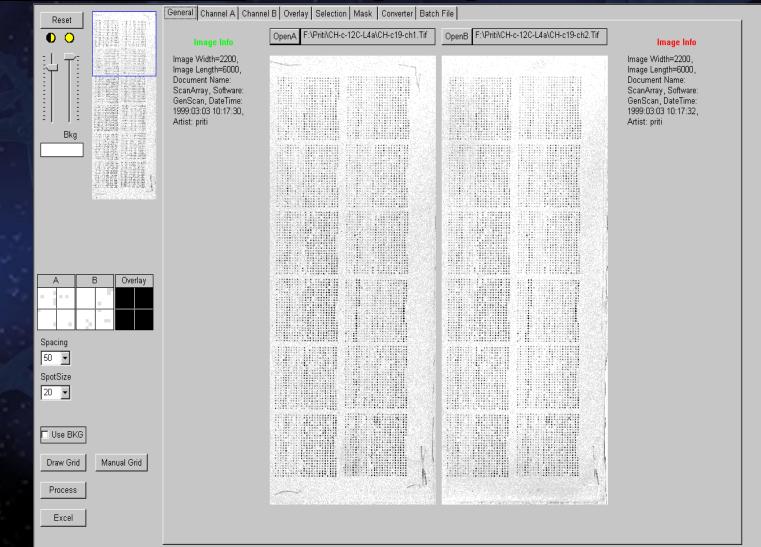
Reproducibility

Measure - median vs. mean (integrated intensity)

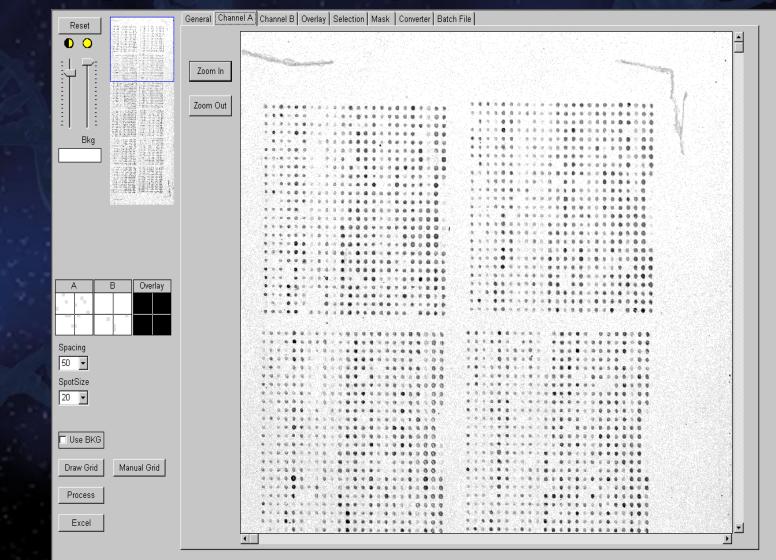
Quality measures



TIGR Spotfinder Loading Image Data



TIGR Spotfinder Zooming In

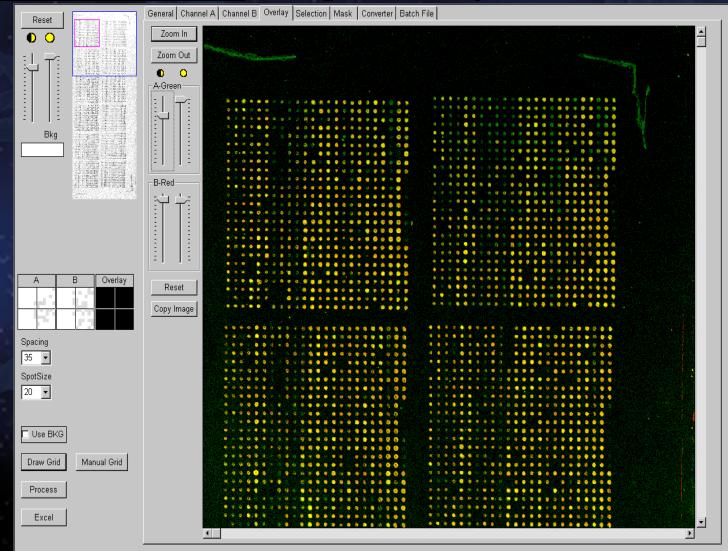


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TIGR Spotfinder Image Overlay



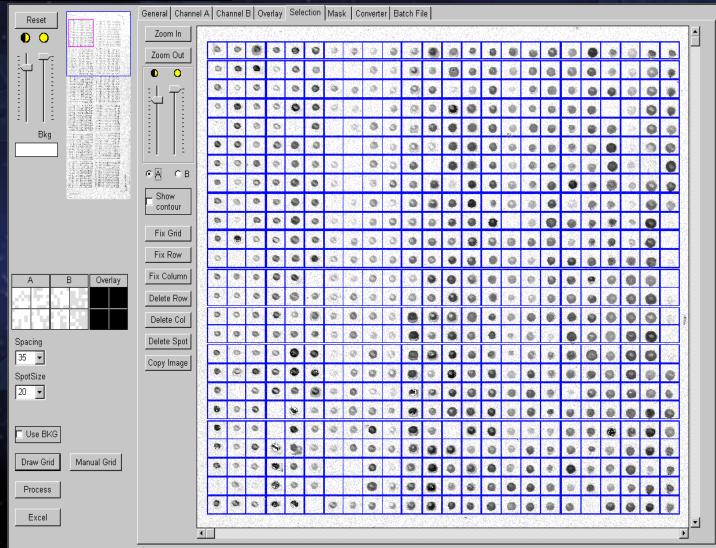
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TIGR Spotfinder Region Selection

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TIGR Spotfinder Grid Determination



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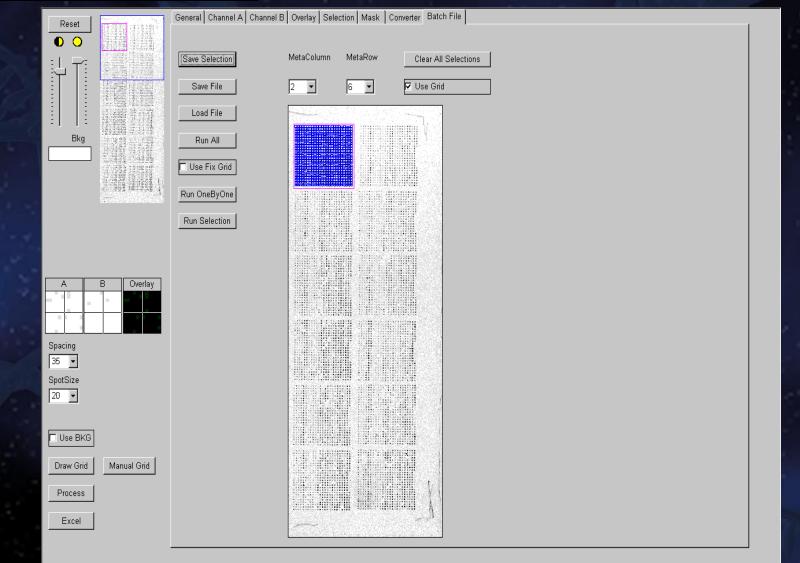
TIGR Spotfinder Grid Adjustment

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TIGR Spotfinder Spot Determination

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TIGR Spotfinder Batch Mode



<u>TIGR Spotfinder</u> Data Ouptut to Spreadsheet

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1	0.6583662	0.3200546	0	1.4709515	5.5659516	0.8221982	0	1.2417782	15.806612	0.6525556	2.2853259	1.1422822	1.5317368	0.5656727	0.2204492	0.6117383	0.3719255	
2	0.7558224	0.4726607	0.6765916	2.1505848	2.4587546	0	16.717262	0	0.545069	0.2482868	0.6166225	0.6360926	2.7529202	0.4951814	0.7414343	0.7803267	0.3516873	C
3	0.6014479	0.6786517	0.4542122	0.8245431	1944	0.9568038	0	1.162365	0	0.4939302	0.5924173	0.4673734	0.7887445	0.6112968	0.6759708	0.3229271	0.4144771	<u> </u>
4	0.7138603	0.627963	0.6439672	0.7704063	0.8925518	0	0.4864845	0	1.5022185	1.4716133	0.6375733	1.8214447	1.4876398	0.5303082	0.9627375	0.5498852	0.4752789	<u> </u>
5	19867	0.5250862	0.5794229	4.6387927	1.6258561	0.6763154	0.6856697	0.6019967	0.9303186	0.7640353	4732	0.1956629	0.6116118	0.7179828	0.511428	0.5777325	0.3184048	
6	0.5597972	0.573433	0.5087411	1.0288527	0.8213453	1.088831	0.5182413	7.3380015	0.5439334	0.7649303	0.6407183	0.8737587	2.5823746	3.0950537	1.0833581	0.5501509	1.4692662	
7	0.8181596	5.0143941	0.4104919	0.8590075	2.4741482	0.511308	0.2833126	0.7497842	0.5266239	0.5785949	0.5328029	0.5844418	1.3443897	0.6685372	0.7394423	0.4709574	0.3382549	<u> </u>
8 9	0.4960111	0.9603443	0.6195294	0.8830633	1.8163842	0.7538587	3.0679385 0	0.888154	0.8022219	0.5235721	0.4453822	0.6102882	4.3070269	0.5359387	1.1485683	0.5624667	0.5095583	
9 10		0.331194	0.9343573	0.4383423	0.5945796	2.1851852	2.2649694	1.5584225	0.6857057	0.3514542	0.9283824	1.5142216	0.5062739	2.3805146	0.6122436	0.4766529	1.1163227	<u>_</u>
10 11	0.667834	0.6553847	0.628634	0.5791472	0.6932317	4.3197004		0.8664637	0.4157382	13935 0.2044601	2.2690295	0.7714797 18592	0.4356746	1.8112027 0.805083	0.4992847	0.3741092	0.5494838	-
11 12	0.377371	0.6553847	0.6047932	0.6122858	27360	1.0413585 22.128571	2.6687442	0.6999262	1.1087549	0.2044601	0.7793285	28160	0.3790219	41403	0.5479772	0.71932	0.6865773	_L
12 13	0.3941197	0.4684356	0.549851	0.3378463	0.5800944	0.4016139	0.4345092	0.6999262	0.5058177	0.193656	0.6716211	28160	0.3814872	0.4716132	0.3442173	0.71932	0.8772932	
14	0.6270496	0.6417714	0.6545356	0.5588574	0.6905183	0.9231741	0.9723208	0.6572631	0.5058177	0.2494581	0.0710211	0.6706132	0.5712125	43.367568	0.3442173	0.0011077	0.602618	<u>_</u>
15	0.9472301	0.4330814	0.5178506	0.5566574	0.6018318	0.5159809	0.9723208	0.3768859	0.5054452	1.0501037	0.6222883	0.5850231	0.6007415	43.367560	0.7071647	1.6884949	0.9790521	
16	0.6911812	0.4346258	1.1916842	0.4309419	8.9242692	43848	0.04013	96.033457	0.6009259	0.7480124	0.6310674	32015	0.6647776	0.6107295	0.4037619	0.4548992	0.5031051	<u>_</u>
17	1.9877921	0.8708843	0.5843845	0.4303413	10239	0.630209	0.8887241	30.033437 0	0.5655554	0.924516	1.1751844	0.9742245	0.5055666	0.675697	0.6563043	0.6590467	0.7881861	-
18	0.8784821	0.762783	0.6200871	0.4537044	0.8391829	0.4867697	0.4590029	15229	3.3292708	0.6948884	0.6198393	0.951506	0.9830806	0.5503178	0.297055	0.5330157	0.6457372	<u> </u>
19	0.7443983	0.3993024	0.2824506	9,7648983	1.6374832	0.4936004	4.3733205	5,9774657	3.660936	0.9652109	0.7294435	1.0556538	0.2780486	0.5547685	0.4831949	0.6634127	0.8785753	
20	0.6150365	0.7199398	0.3694191	3.0180843	17.80643	0.6281572	45.43388	2.4153739	20010	2.0190544	0.7906932	0.5755896	23672	21.117735	47.738657	3.7311394	0.561419	-
21	0.3831165	0	0.7697642	0	0	0.9106237	0.559151	0.4493976	0.5317561	1.9017256	0.454886	0.6134742	0.4145606	0.4072714	0.6959529	0.5330013	0.5375891	C
22	0.7813621	0.9994261	0.6439033	0.4590316	3.3381008	0.4194966	0.661315	0.6708735	1.2265628	6.7601277	0.8665072	0.4766182	0.6210309	0.2603647	0.4316437	0.6079227	0.6760341	-
23	0.9928715	0.8081178	0.9006058	3.9962708	0.5796124	0.326307	0.5219831	1.2754642	13366	0.7130079	0.6032611	0.4392349	0.2545188	1.0666558	0.4684213	0.7079562	0.4384588	2
24	0.6423902	0.5016539	0.6781809	0	0	0.4935037	0.3806448	0.4131515	0	1.7617517	0.608026	0.5602693	1.2712959	0.3531592	0.8132697	0.5016375	0.5417107	
25	2.7186579	0.5461783	4.7906256	1.3970144	1.2894225	33492	0.5698123	0.6633413	45537	7.7535278	0.4481022	0.8431524	0.5930069	0.5466802	0.9798421	0.5280782	0.6872405	C
26	0.4448915	0.2928477	1.138561	0.872978	2.9418556	3.2201626	0.4299104	0.7401343	0.8747521	0.6191381	0.5955113	0.7019722	0.7324154	0.3111703	0.6549464	0	0.5135729	1
27	0.8660603	0.4925402	0.5055647	18821	0	0	0.4171291	0.4299915	0.5592586	0.367861	0.5143132	0.95747	0.6374241	0.5562525	0	0.6587389	0.6611734	C
28	0.6870497	0.5846247	0.5644515	5.7504524	0.7351635	2.645988	0.2871387	0.6415056	28739	0.6660191	0.5158808	0.9255612	0.5047607	0.6047711	0.6464061	0.7697136	0.7831176	C
29	0.8289175	0.5033416	0.5644581	5.5740416	0.6687968	1.2334758	0	0.6071581	0.5320682	0.9103183	0.4253956	2.0644427	0.3088956	0.6550209	2.8981733	0.3853836	0.6782611	C
30	0.6018762	0.7041271	0.8834399	0.5162058	0.5014956	3.0018989	0.4932798	1.2487259	0.4648862	1.0122426	0.3936311	0.5598506	0.2981063	0.5395712	1.1278056	0.5936225	1.2177372	1
31	1.8010817	0.6359243	0.5820145	1.2524697	1.64248	19.682836	0.6768014	0	0.8281743	0.788474	0	0.9617461	0.468756	0.7439989	0.4900083	0.6258416	0.5829106	C
32	0.5271594	0.788637	0.8683437	0.4918654	0.8378943	28691	0.3787699	0.697312	0	1.0318284	7.5672938	0.8437288	1.162031	0.529426	0.2146548	0.5499871	0.3565854	С
33	1.1121659	0.7409447	0.6622219	0.7243704	0.4079886	2.7667886	1.5015783	0.5296397	0.4397992	0.8804963	0.4392882	0	0.6234124	0.7472773	0.3598265	0.6405565	0.9221139	С
34	0.6449821	0.8512406	0.4972888	3.7825806	0.6082631	0.5507858	0.9195138	0.8752735	0.664297	0.5932464	0.3333748	0.6606828	0.71511	0.6435152	0.4954949	0.6247832	0.6587191	C
35	28964	0.9977072	0.5916031	5.875817	2.1456126	24.453188	0	0	0.4265595	0.7368994	2.5250656	0.7106283	0.5794094	0.6161443	0.57432	0.4170127	0.9118404	C
36	1.1047913	1.1000884	0.7202179	0	1.0189481	1.0768023	0.6247294	6.124451	8.1567366	0.5836243	0.611794	3.1444078	0.6042322	18.418329	0.4720763	0.5798694	1.1561933	1
37	0	0.7718797	0.91057	0.9746901	0.6604389	0.6350821	0.6017641	0.729244	0.5387656	2.3600445	0.6795764	0.7348712	0.3155152	0.470978	0.4544045	0.5517463	0.3706317	C
38 0.7383087 0.7188266 1.1767919 1.2322487 0.8287257 0.0.4339617 0.8561166 0.3670076 0.599923 0.873193 23288 0.6733388 0.4585579 0.5989658 0.4193539 0.3398962 ▼																		
Integral 1 / Integral 2 / Integral Ratio / Spot Area / Saturation factor / Image: Spot Area / Saturation factor / Ready Image: Spot Area / Saturation factor /																		
Re	Click here to	begin 0	Y	': 0	pixe	l value in	A: 12	р	ixel value	in B : 47								
		box - Microso	ift Outlook	TIGR_S	Spotfinder - 0.0	0.0	Microsoft E:	cel - Book	1							stor 4	 6:3	8 PM
_	<u> </u>		_								_		-			- Internet w		_

Output includes: Integrated Intensity 1, Integrated Intensity 2, Ratio, Spot Area, Saturation, Mean and Median Intensities, Quality Factors

<u>Comparison of Mean, Median, and Mode Ratios</u>

1.000

1.067

1.008

1.098

0.980

1.04

1.030

1.013

0.896

1.026

1.00

0.977

1.06

1.136

0.929

0.877

1.009

0.068

averade

stdev

Mean ratio

1.012	0	0.966	0.987	0.897	
1.135	0	1.037	1.034	1.015	
1.008	0	1.058	1.008	1.058	
1.079	0	1.059	1.061	1.026	
1.022	0	1.069	1.031	1.019	
1.070	0	1.032	1.024	1.139	
0.986	0	1.058	1.064	1.047	
1.057	0	0.990	1.063	1.022	
0.935	0	1.105	1.069	1.079	
1.094	0	1.024	1.057	0.892	
1.014	0	1.040	0.997	1.019	
0.985	0	1.005	1.067	1.035	
1.011	0	1.033	1.035	1.143	
1.232	0	0.996	1.169	1.077	
0.819	0	1.085	1.118	1.039	
0.942	0	0.999	1.129	1.061	
1.025		1.035	1.057	1.036	
0.092		0.037	0.049	0.067	

averade

stdev

Median ratio

0.930

1.053

1.042

1.047

0.998

1.040

1.081

0.987

1.025

1.028

1.067

0.928

1.105

0.963

1.033

0.974

1.019

0.051

1.053

1.056

1.056

1.034

1.004

1.045

1.029

1.085

1.007

1.072

1.020

1.094

1.027

1.067

1.083

1.133

1.054

0.034

0.898

1.015

1.003

1.026

0.955

1.074

1.134

1.014

1.079

0.874

1.014

0.979

1.096

1.020

1.062

1.110

1.022

0.072

Mode ratio

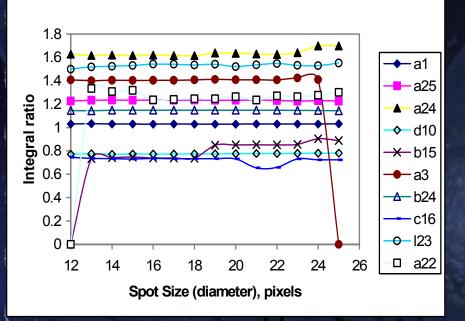
	1.398	0	1.113	0.830	0.518
	1.984	0	0.554	0.906	5.721
	1.536	0	1.113	1.152	0.570
	1.051	0	2.490	1.684	1.437
	2.794	0	0.976	1.544	1.651
	1.095	0	1.564	1.203	1.516
	1.332	0	1.253	0.614	94.797
	1.697	0	16.921	2.039	0.788
	0.550	0	1.065	0.873	0.916
	1.022	0	0.742	0.377	0.681
	1.707	0	1.714	1.411	2.167
	1.784	0	0.392	0.976	1.080
	2.269	0	0.318	1.708	0.615
	1.932	0	0.604	1.331	0.579
	0.052	0	1.618	0.380	1.254
	0.641	0	0.543	1.373	1.022
average	1.428		2.061	1.150	7.207
stdev	0.690		4.004	0.479	23.391

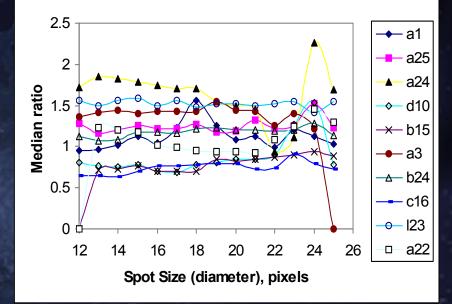
A comparison of Cy3/Cy5 ratios calculated using the mean, median, and mode ratios for control spots that should have a measured ratio of 1 for the 1st, 3rd, 4th, 5th columns.

<u>Integral (Mean) Ratio vs. Median Ratio</u>

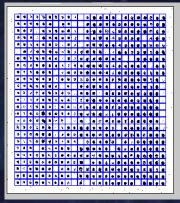
Integral Ratio vs. used Spot Size

Median ratio vs. used Spot Size





A comparison of Cy3/Cy5 ratios for various spot sizes using either the integrated intensity or the pixel median. In this case, the actual spot size is approximately 15 pixels in diameter.



Microarray Expression Analysis

Species Selection

Gene

Differential Growth – Conditions

RNA Preparation and Labeling

Competitive Hybridization

Spot on a – <u>Slide</u>

Fluoresence Intensity **Expression Measurement**

Data Analysis Issues

• Presentation

Multiple Views

Normalization

Identification of Differentially Expressed Genes

Multiple Experiments

Why Normalize Data?

Goal is to measure ratios of gene expression levels $(ratio)_i = R_i/G_i$ where R_i/G_i are, respectively, the measured intensities for the *i*th spot.

In a self-self hybridization, we would expect all ratios to be equal to one: P/C = 1 for all *i*. But they may not be

 $R_i/G_i = 1$ for all *i*. But they may not be.

Why not?

Unequal labeling efficiencies for Cy3/Cy5

- Noise in the system
- Differential expression

Normalization brings (appropriate) ratios back to one.

Normalization Approaches

Total Intensity

- Linear Regression
- Ratio statistics described by Chen, Dougherty, & Bittner *J. Biomed. Optics* (1997) 2(4) 364-374
 Iterative log(ratio) mean centering

Any of these using:

- Entire Data Set
- User-defined Data Set/Controls

Normalization Approaches Entire Data Set Probe Quantification less important • No assumption on which genes constitute "housekeeping" set Uses all the data • No independent confirmation **User-defined Data Set/Controls** • Requires definition of "housekeeping" set or good added controls Requires good RNA quantitation **Ignores much data** THE INSTITUTE FOR GENOMIC RESEARCH

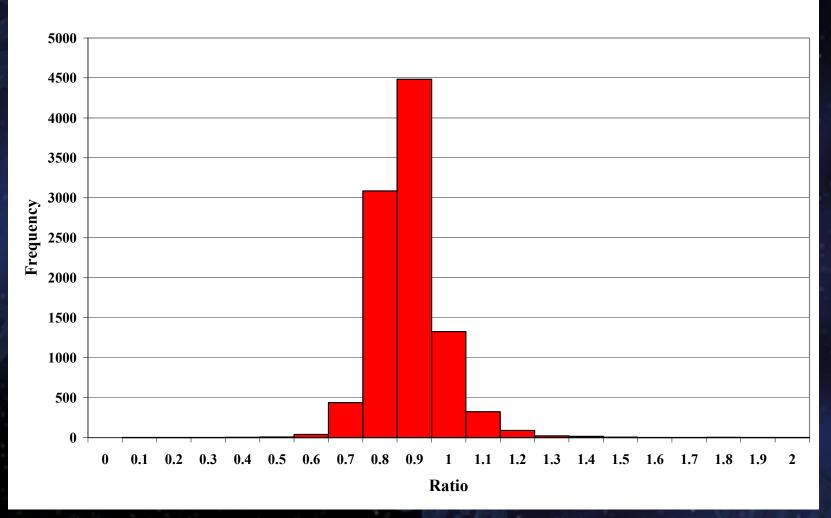
Normalization Approaches

Solution(?)

Experiment dependent
Use a combination of techniques
SMART Experimental design

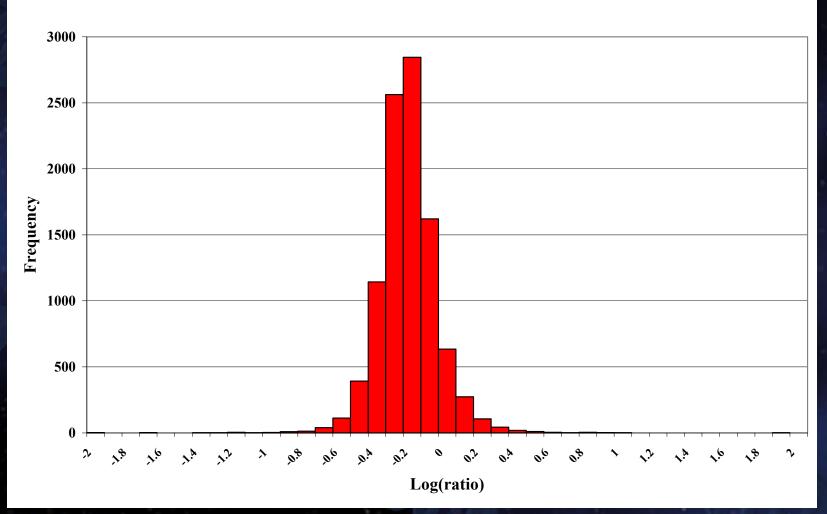


Ratio Histogram



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Log(ratio) Histogram



Normalization Approaches: Total Intensity

Assumption: Total RNA (mass) used is same for both samples.

So, averaged across thousands of genes, total hybridization should be the same

$$V = \frac{\sum_{k=1}^{Narray} R_k}{\sum_{k=1}^{Narray} G_k}$$

Normalization: $G'_k = NG_k$ and $R'_k = R_k$.

Normalization Approaches: Linear Regression

Assumption: Total RNA used is constant, some genes expressed with ratio of 1, slope of best fit line normalized to 1

Normalization Factor:

$$R_k = \beta_0 + \beta_1 G_k + u_k$$

$$S(\beta_0, \beta_1) = \sum_{k=1}^n \mu_k^2 = \sum_{i \setminus k=1}^n (R_k - \beta_0 - \beta_1 G_k)^2$$

The values of β_0 and β_1 that minimize $S(\beta_0, \beta_1)$, b_0 and b_1 , are given by

$$b_{1} = \frac{\sum_{k=1}^{n} (R_{k} - \overline{R})(G_{k} - \overline{G})}{\sum_{k=1}^{n} (G_{k} - \overline{G})^{2}} \quad \text{and} \quad b_{0} = \overline{R} - b_{1}\overline{G}$$
where $\overline{R} = \frac{\sum R_{k}}{n}$ and $\overline{G} = \frac{\sum G_{k}}{n}$.

Normalization:
$$G'_k = \left[\frac{1}{b_1}\right] G_k$$
 and $R'_k = R_k$.

Sormalization Approaches: Ratio Statistics (1)

Assumption: Total RNA used is constant, some genes expressed with ratio of 1, variations are functions of the common mean

 $\sigma_{G_k} = c\mu_{G_k}$ and $\sigma_{R_k} = c\mu_{R_k}$, with $\mu_{G_k} = \mu_{R_k} = \mu_k$.

Probablilty Density for Ratio
$$T_k$$
: $f_{T_k}(t) \approx \frac{(1+t)\sqrt{1+t^2}}{c(1+t^2)\sqrt{2\pi}} \exp\left[\frac{-(t-1)^2}{2c(1+t^2)}\right]$

This density can be used to calculate the mean, standard deviation and confidence interval limits for the distribution of measured ratio values. As functions of c, these parameters can be estimated using a polynomial approximation

$$y = a_3c^3 + a_2^2c^2 + a_1c + a_0$$

with constants are chosen appropriately:

 $\mu: \quad (a_3, a_2, a_1, a_0) = (0.364, 1.279, -0.0427, 1.001)$ $\sigma: \quad (a_3, a_2, a_1, a_0) = (-2.805, 2.911, -2.706, 0.979)$ lower limit at 95% confidence: $(a_3, a_2, a_1, a_0) = (28.644, -2.830, 3.082, 0.989)$

upper limit at 95% confidence: $(a_3, a_2, a_1, a_0) = (-5.002, .4.462, -3.496, 0.9968)$

Normalization Approaches: Ratio Statistics (2)

Assume that the population mean $\mu_0 = 1$ and let the first approximation of the normalization parameter m₁ be equal to the calculated sample A first approximation of *c*, \hat{c}_1 , is calculated using

$$\hat{c}_{i} = \left[\frac{1}{n}\sum_{j=1}^{n}\frac{(t_{j}-1)^{2}}{(1+t_{j}^{2})}\right]^{1/2}$$

where the sum is over the n elements taken initially between the one-half and twice the sample mean.

Upper and lower limits at the 95% confidence level, θ_1 and θ_2 , are then calculated using \hat{c}_1 and the previous approximation.

A normalization factor \hat{m}_1 is calculated using

$$\hat{m}_i = \frac{1}{\hat{\mu}_{i-1}} \left(\frac{1}{n} \sum_{j=1}^n t_j \right),$$

where, again, we take $\hat{\mu}_0 = 1$, the sum is over the *n* array elements used to estimate \hat{c}_1 , and *i* is an index used to count the number of iterations

The individual ratios are then rescaled using

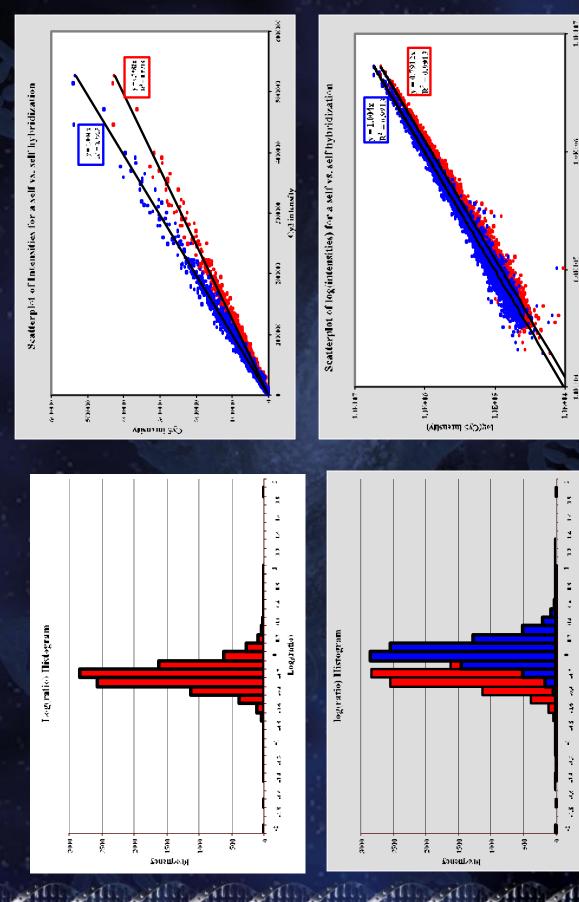
$$t'_{k} = \frac{t_{k}}{\hat{m}_{i}} = \frac{R_{k}}{(\hat{m}_{i}G_{k})} = \frac{R'_{k}}{G'_{k}}.$$

This process is then iterated until the calculated value of the mean estimator converges to a fixed value.

The upper and lower confidence limits for the normalized experimental data are then calculated as

$$\theta_1' = \hat{m}\theta_1$$
 and $\theta_2' = \hat{m}\theta_2$

and (θ'_1, θ'_2) are used to define the limits for identification of differentially expressed genes

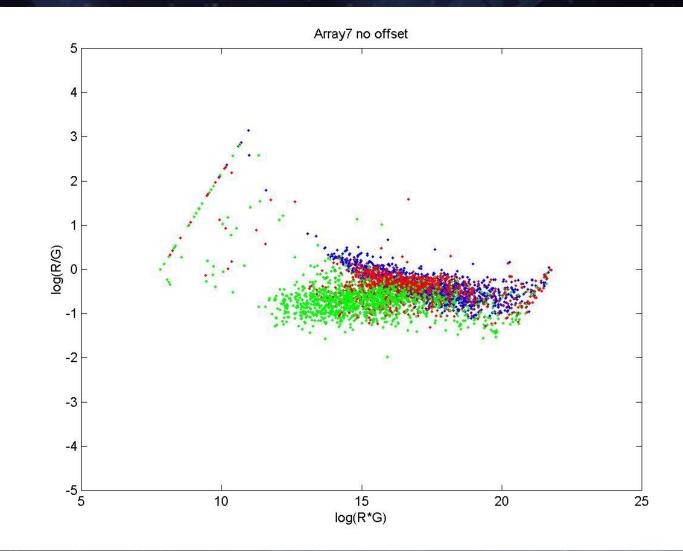


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log(Cy3 Intensity)

(effert)_e gel

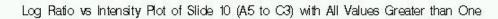
Bad Data from Parts Unknown

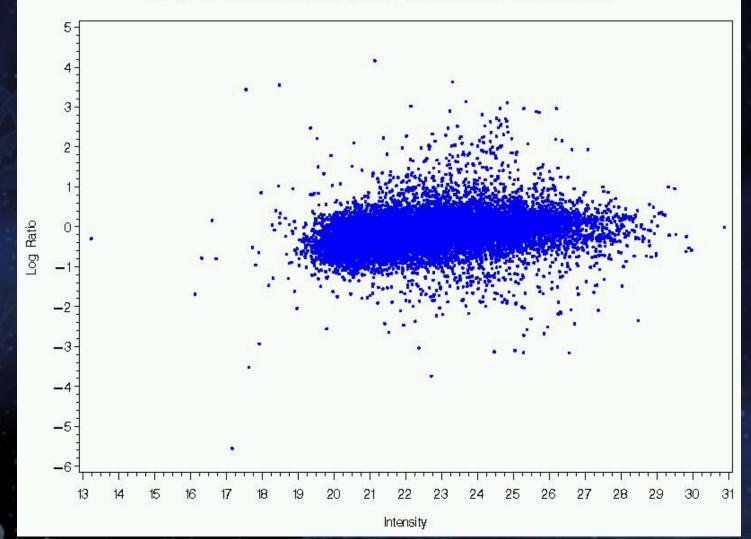


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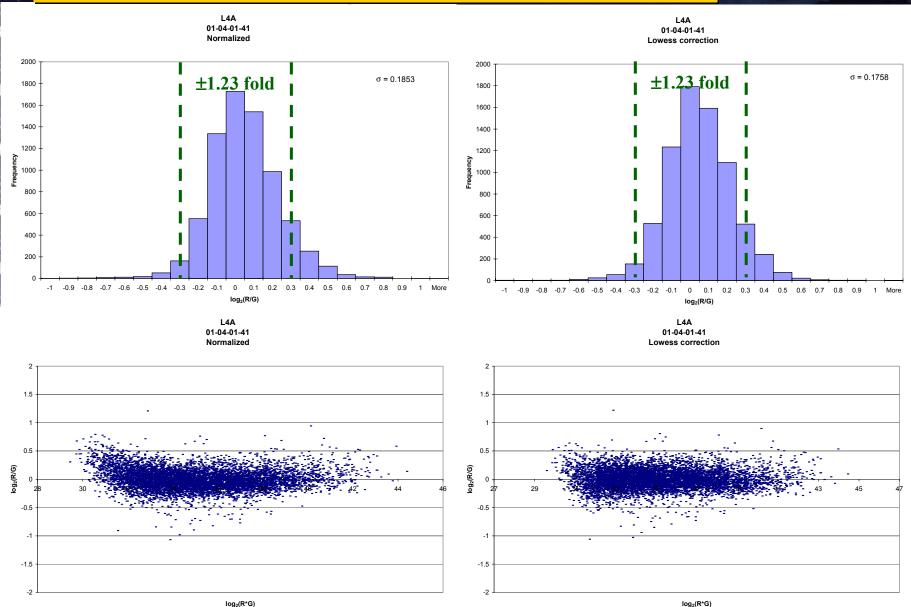
Gary Churchill

Good Data from TREX





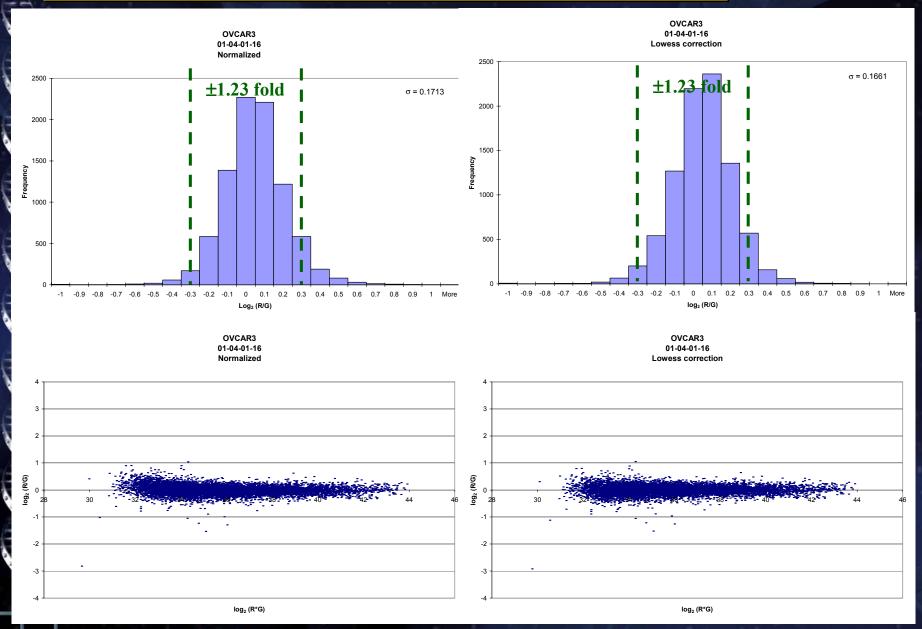
Normalization using local linear regression



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Ivana Yang, John Quackenbush

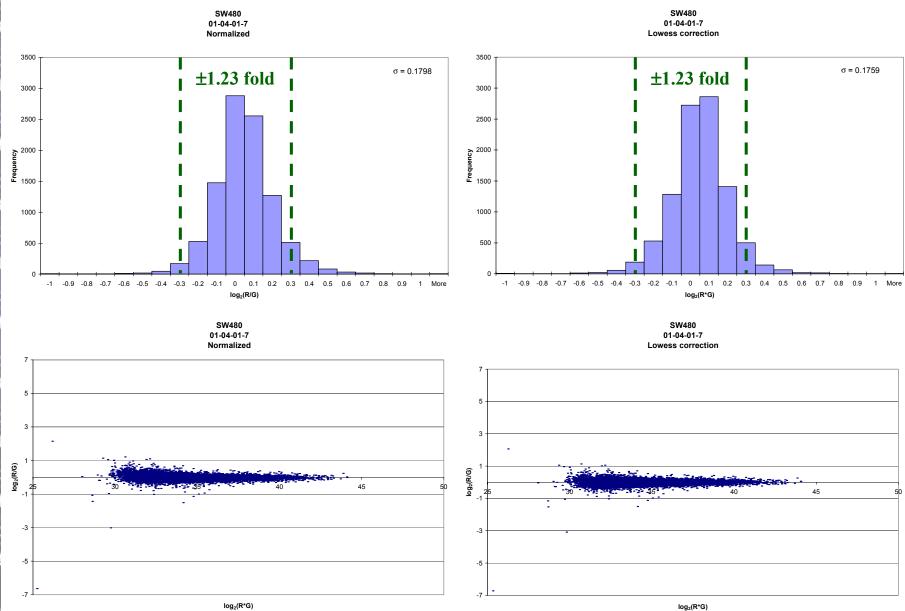
Normalization using local linear regression



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Normalization using local linear regression



log₂(R*G)

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Multiple Experiments?

Goal is identify genes (or experiments) which have "similar" patterns of expression

This is a problem in data mining

"Clustering Algorithms" are most widely used

Types

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- Agglomerative: Hierarchical
- Divisive: k-means, SOMs
- Others: Principal Component Analysis (PCA)

All depend on how one measures distance

Expression Vectors

Crucial concept for understanding clustering

Each gene is represented by a vector where coordinates are its values log(ratio) in each experiment

- $x = \log(ratio)_{expt1}$
- $y = \log(ratio)_{expt2}$
- $z = \log(ratio)_{expt3}$

etc.

Expression Matrix

These gene expression vectors of log(ratio) values can be used to construct an expression matrix

		t	(† 3)			t
	Expt	Expt	xpt	xpt	xpt	xpt
Gene ₁	-1.2	-0.5	0	0.25	0.75	1.4
Gene ₂	0.2	-0.5	1.2	-0.25	-1.0	1.5
Gene ₃	1.2	0.5	0	-0.25	-0.75	-1.4
etc.	Y					

This is often represented as a red/green colored matrix



Distance metrics

Distances are measured "between" expression vectors

Distance metrics define the way we measure distances

Many different ways to measure distance:

- Euclidean distance
- Pearson correlation coefficient(s)
- Manhattan distance
- Mutual information
- Kendall's Tau
- 🔷 etc.

Each has different properties and can reveal different features of the data

Distance Matrix

Once a distance metric has been selected, the starting point for all clustering methods is a "distance matrix"

	Gene	Gene ₂	Gene ₃	Gene ₄	Genes	Gene
Gene ₁	0	1.5	1.2	0.25	0.75	1.4
Gene ₂	1.5	0	1.3	0.55	2.0	1.5
Gene ₃	1.2	1.3	0	1.3	0.75	0.3
Gene ₄	0.25	0.55	1.3	0	0.25	0.4
Gene ₅	0.75	2.0	0.75	0.25	0	1.2
Gene ₆	1.4	1.5	0.3	0.4	1.2	0

The elements of this matrix are the pair-wise distances. Note that the matrix is symmetric about the diagonal. TIGR THE INSTITUTE FOR GENOMIC RESEARCH

Hierarchical clustering

Select the data you want to cluster

"Filter" (normalize) the data appropriately and select distance

Apply method:

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- 1. Search through the distance matrix and find the two most similar clusters. This is the first true stage in the "clustering" process. If several pairs share the same similarity, use a predetermined rule to decide between alternatives.
- 2. Fuse the two selected clusters to produce a new cluster that now contains at least two objects.
- **3.** Calculate the distances between this new cluster and all other clusters. There is no need to calculate *all* distances since only those involving the new cluster have changed.

Repeat steps 1-3 until all objects are in one cluster.

<u>k-means clustering</u>

Select the data you want to cluster and filter; select distance

Apply method:

the second second

- **1.** All initial objects are randomly assigned to one of k clusters (where k is an input parameter to the algorithm).
- 2. An average expression vector is then calculated for each cluster and this is used to compute the distances between clusters.
- **3.** Objects are moved between clusters and intra- and inter-cluster distances are measured with each move. Objects are allowed to remain in the new cluster only if they are closer to it than to their previous cluster.
- 4. Following each move, the expression vectors for each cluster are recalculated.
- **5.** The shuffling proceeds until moving any more objects would make the clusters more variable.

Self Organizing Maps (SOMs)

Select the data you want to cluster and filter; select distance

Apply method:

- **1.** Random vectors are constructed and assigned to each partition. (where the number and geometry are input parameters).
- 2. A gene is picked at random and using a selected distance metric, the reference vector that it is closest to the gene's is identified .
- **3.** The reference vector is then adjusted so that it is more similar to the randomly picked gene's. The reference vectors that are nearby on the two dimensional grid are also adjusted so that they too are more similar to the randomly selected gene .
- 4. Steps 2 and 3 are iterated several thousand times, decreasing the amount by which the reference vectors are adjusted and increasing the stringency used to define closeness in each step. As the process continues, the reference vectors are converge to fixed values.
- **5.** Finally, the genes are mapped to the relevant partitions depending on the reference vector to which they are most similar.

Principal Component Analysis (PCA)

Select the data you want to cluster and filter

Apply method: OK, this gets a bit complicated....

Basically:

AN THE ARE

- **1.** We find the eigenvectors of the expression matrix
- 2. We select those with the greatest eigenvalues
- **3.** We project our data on the eigenvectors with the three greatest eigenvalues
- 4. And make pretty pictures

Support Vector Machines (SVM)

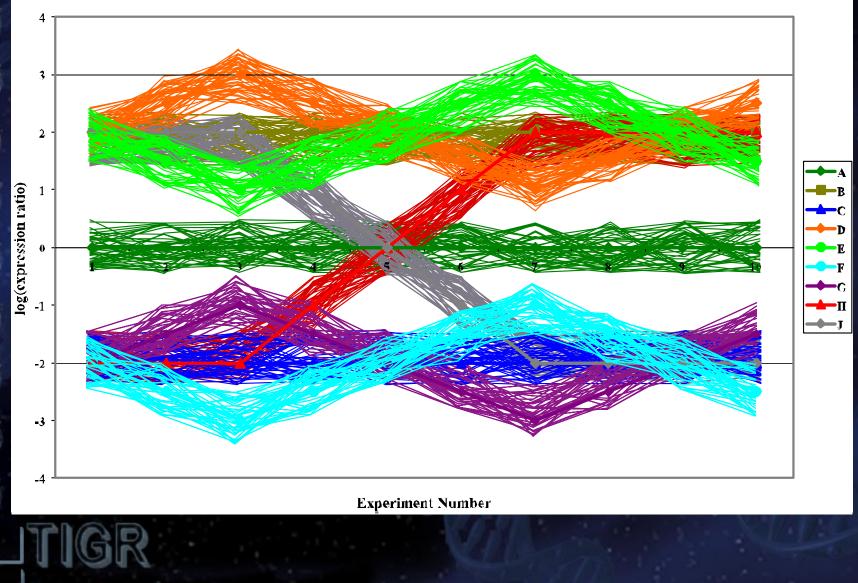
Select the data you want to cluster and filter

Apply method: OK, this gets even more complicated....

Basically this is a neural network approach to finding dividing your data into genes "like" and "unlike" a training set....
1. Pick a set of genes you are know about (your training set)
2. Train the SVM. This produces a pattern that can be recognized
3. Screen the data using the SVM model

TIGR MeV: Test Data Set

Gene Expression Families



Hierarchical Clustering

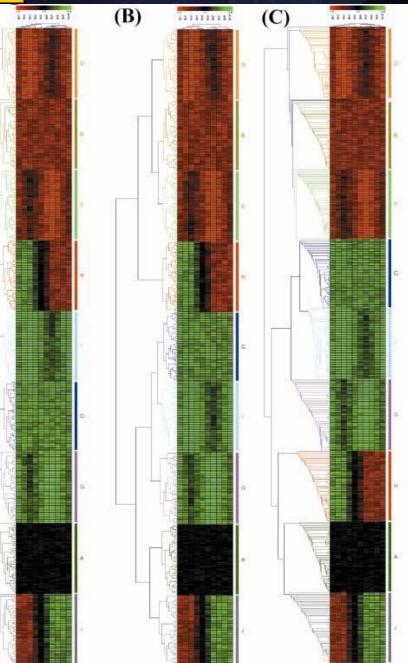
(A)

(A) Average Linkage

(B) <u>Complete Linkage</u>

(C) Single Linkage

Even related algorithms produce slightly different views of the data.



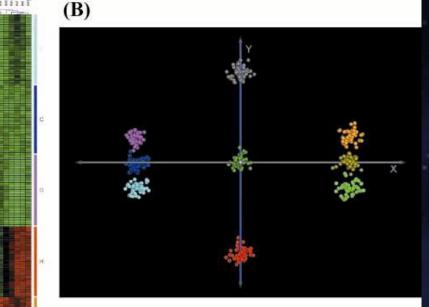
Hierarchical Clustering and PCA

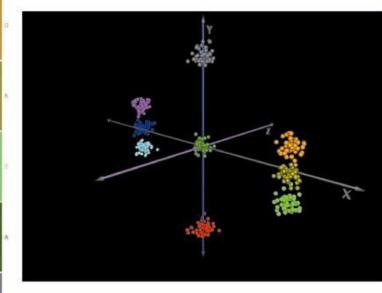
(A)

(A) Average Linkage

(B) <u>PCA</u>

Separate clusters may have more or less support when using different algorithms.

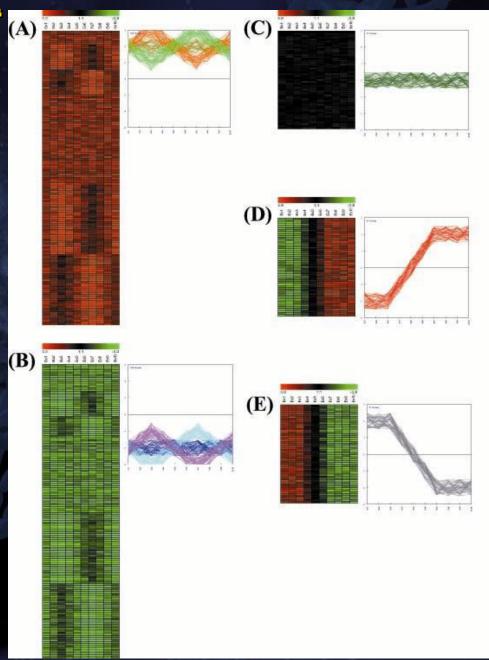




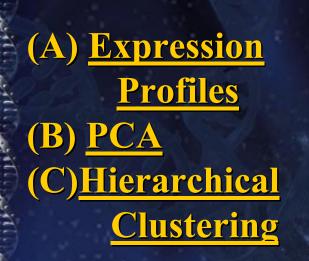
k-means Clustering

Separate clusters may have more or less support when using different algorithms.

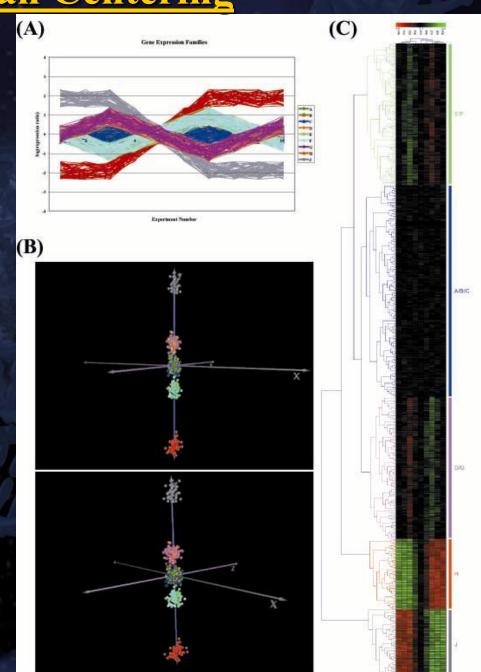
Note colors are based on hierarchical clustering Results.



The effects on Mean Centering



Adjusting the data can have profound effects, but allow different patterns to be seen.



Very Useful Microarray URLs

Leming Shi TIGR MGED Wentian Li EBI Terry Speed Joe Derisi Pat Brown NCGR Stanford

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HAPI

http://www.gene-chips.com http://pga.tigr.org/tools http://www.mged.org http://linkage.rockefeller.edu/wli/microarray http://industry.ebi.ac.uk/~alan/MicroArray http://stat-www.berkeley.edu/users/terry/zarray/Html http://stat-www.berkeley.edu/users/terry/zarray/Html http://www.microarrays.org/index.html http://cmgm.stanford.edu/pbrown/mguide/ http://www.ncgr.org/research/genex/other_tools.html http://www.dnachip.org

http://array.ucsd.edu

Acknowledgments

The TIGR Gene Index Team

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